Bromopyrrole Alkaloids from Marine Sponges of the Genus Agelas[⊥]

Tetsuro Yasuda,[†] Atsushi Araki,[†] Takaaki Kubota,[†] Junji Ito,[§] Yuzuru Mikami,[§] Jane Fromont,[‡] and Jun'ichi Kobayashi^{*,†}

Graduate School of Pharmaceutical Sciences, Hokkaido University, Sapporo 060-0812, Japan, Medical Mycology Research Center, Chiba University, Chiba 260-0856, Japan, and Western Australian Museum, Locked Bag 49, Welshpool DC, WA 6986, Australia

Received October 15, 2008

Four new bromopyrrole alkaloids, nagelamides O (1) and P (2) and mukanadins E (3) and F (4), were isolated from three collections of Okinawan marine sponges of the genus *Agelas*. The structures and stereochemistry of 1-4 were elucidated on the basis of their spectroscopic data.

Bromopyrrole alkaloids are known to be one of the most common metabolites contained in marine sponges.¹ During our search for bioactive substances from Okinawan marine organisms,² we have previously isolated several bromopyrrole alkaloids with unique cyclic skeletons from sponges of *Agelas* species.^{3,4} Recently, we have investigated extracts of three collections of Okinawan marine sponges of the genus *Agelas* and have isolated two new dimeric bromopyrrole alkaloids, nagelamides O (1) and P (2), and two new monomeric bromopyrrole alkaloids, mukanadins E (3) and F (4). Herein, we describe the isolation and structure elucidation of 1-4.

The sponge *Agelas* sp. (SS-983) collected off Seragaki, Okinawa, was extracted with MeOH, and the MeOH extract was partitioned between EtOAc and H₂O. *n*-BuOH-soluble materials of the aqueous layer were subjected to passage over an LH-20 column and a C₁₈ column, followed by C₁₈ HPLC, to yield nagelamides O (1, 0.0014%, wet weight) and P (2, 0.0005%). The other two collections of Okinawan sponges of *Agelas* species (SS-1056 and SS-1077) were obtained off Gesashi and off Unten-Port, respectively. Mukanadin E (3, 0.00032%) was isolated from EtOAc-soluble materials from the MeOH extract of the sponge of *Agelas* species (SS-1056) collected off Gesashi, Okinawa, while mukanadin F (4, 0.00094%) was obtained from that of the sponge of *Agelas* species (SS-1077) collected off Unten-Port, Okinawa, by the same procedure.



The ESIMS of nagelamide O (1) showed pseudomolecular ion peaks at m/z 765, 767, 769, 771, and 773 (3:10:12:6:1), indicating the presence of three bromine atoms and one chlorine atom in the molecule. Nagelamide O (1) was revealed as possessing the

molecular formula C₂₂H₂₄Br₃ClN₁₀O₄ by HRESIMS [m/z 764.9315 $(M + H)^+$, $\Delta + 1.6$ mmu]. The UV absorption [λ_{max} 275 (ϵ 9400) nm] was attributed to a substituted pyrrole chromophore, while IR absorptions indicated the occurrence of OH and/or NH (3450 cm⁻¹) and amide carbonyl (1680 cm⁻¹) functionalities. The ¹H NMR (Table 1) spectrum showed 12 D₂O-exchangeable signals ($\delta_{\rm H}$ 12.71, 11.82, 10.19, 9.92, 9.27, 9.25, 8.87, 8.62 (2H), 8.34, 8.21, 7.82, and 7.53). The ¹³C NMR (Table 1) spectrum disclosed 22 signals due to two amide carbonyl carbons, seven sp² quaternary carbons, three sp² methines, two sp³ quaternary carbons, six sp³ methines, and two sp³ methylenes. Among the ¹³C NMR signals of 1, two amide carbonyl carbons [$\delta_{\rm C}$ 160.1 (2C)], five sp² quaternary carbons $(\delta_{\rm C} 127.8, 126.8, 104.8, 98.1, \text{ and } 95.1)$, and three sp² methines (δ_C 121.4, 113.5, and 111.6) were ascribed to di- and monobromopyrrole carbonyl moieties (N-1-C-6 and N-1'-C-6') by comparison with those of known bromopyrrole alkaloids.^{3,4}

Detailed analysis of the ¹H-¹H COSY and TOCSY spectra of 1 disclosed connectivities from NH-7 to H-10, NH-7' to H-10', H-9 to H-9', NH-1' to H-2', and H-15 to OH-15. HMBC correlations for H-10 to C-11 and C-15, H-15 to C-13, NH-14 to C-11, H-10' to C-11 and C-11', 11'-OH to C-10', NH-12' to C-13', NH-14' to C-11', and H-15' to C-11' and C-13' indicated the presence of a tetracyclic core⁵ (C-9 to C-11, N-12, C-13, N-14, C-15; C-9' to C-11', N-12', C-13', N-14', C-15'). ROESY cross-peaks of H-4/ NH-7 and H-4'/NH-7' implied connectivities of a dibromopyrrole ring to N-7 and a monobromopyrrole ring to N-7' through an amide bond, respectively (Figure 1). ROESY correlations for H-10/H-15', H-15'/11'-OH, and 11'-OH/H-9' indicated α -orientations of H-10, H-9', H-15', and 11'-OH, while β -orientations of H-10, 15-OH, and H-10' were deduced from ROESY correlations for H-10'/ OH-15 and H-10'/NH-12' and a large coupling constant for H-9 and H-10 (J = 12.3 Hz). These results indicated the relative stereochemistry of 1 as shown in Figure 2. Thus, the structure of 1 was assigned as the 2'-debromo form of axinellamine A,⁵ including relative stereochemistry. The absolute configurations at C-9 and C-9' in 1 were assigned as both S from the CD positive exciton split [λ_{max} 281 ($\Delta \epsilon$ +5.2) and 257 ($\Delta \epsilon$ -3.4) nm] due to di- and monobromopyrrole carboxyamide chromophores (Figure 3).⁶ Thus, the structure of nagelamide O was concluded to be 1.

Professor Köck proposed that axinellamine A could be derived from massadine chloride through a hypothetical intermediate preaxinellamine.⁷ Nagelamide O (1) might be generated from massadine chloride or its analogue in a similar manner.

The molecular formula of nagelamide P (**2**) was revealed as $C_{22}H_{20}Br_3N_{10}O_3$ by HRESIMS [*m*/*z* 710.9465 (M + H)⁺, Δ +3.8 mmu]. ¹H and ¹³C NMR data (Table 2) of **2** disclosed the presence of di- and monobromopyrrole carbonyl moieties (N-1–C-6 and N-1'–C-6'). Inspection of the ¹H–¹H COSY and TOCSY spectra of **2** implied connectivities of N-7 to C-10 and N-7' to C-10' (Figure 4). Geometries of the two double bonds at C-9 and C-9' were assigned as both *E* from NOESY correlations for H-8/H-10 and

 $^{^{\}perp}$ Dedicated to Professor David G. I. Kingston of Virginia Polytechnic Institute and State University for his pioneering work on bioactive natural products.

^{*} To whom correspondence should be addressed. Tel: +81-11-706-3239. Fax: +81-11-706-4989. E-mail: jkobay@pharm.hokudai.ac.jp.

[†] Hokkaido University.

[§] Chiba University.

^{*} Western Australian Museum.

position	$\delta_{ m H}$	mult. J (Hz)	$\delta_{ m C}$	position	$\delta_{ m H}$	mult. J (Hz)	$\delta_{ m C}$
1	12.71	brs		1'	11.82	brs	
2			104.8	2'	6.90	brs	121.4
3			98.1	3'			95.1
4	7.00	brs	113.5	4'	6.98	brs	111.6
5			127.8	5'			126.8
6			160.1	6'			160.1
7	8.34	brt,5.7		7'	8.21	brs	
8	3.73	m	37.0	8'	3.44	m	42.4
	3.24	m			3.26	m	
9	1.95	m	49.3	9'	2.61	brs	34.1
10	4.27	d, 12.3	66.0	10'	2.87	brd, 4	51.6
11			83.2	11'			100.0
13			156.0	OH-11'	7.82	S	
NH ₂ -13	9.27	brs		12'	9.92	brs	
	8.87	brs			13'		156.9
14	10.19	brs		NH ₂ -13'	8.62	brs, 2H	
15	5.28	brd, 6.4	78.3	14'	9.25	brs	
OH-15	7.53	brd, 6.4		15'	5.32	brs	79.2

H-8'/H-10' as well as coupling constants for H-9/H-10 (16 Hz) and H-9'/H-10' (16 Hz). HMBC cross-peaks of H-10/C-15' and H-10'/ C-15' indicated connectivities of two imidazole rings at C-11 and C-15', while connectivities of a dibromopyrrole ring to N-7 and a monobromopyrrole ring to N-7' were elucidated from NOESY correlations for H-4/NH-7 and H-4'/NH-7', respectively. Thus, the structure of nagelamide P was elucidated as **2**, which is the 2'debromo form of mauritiamine.⁸ The specific optical rotation, $[\alpha]_D^{22} \sim 0$ (*c* 0.5, MeOH), and the CD spectrum, which was flat between 200 and 400 nm, suggested that **2** was a racemate.⁸

The ESIMS of mukanadin E (**3**) showed pseudomolecular ion peaks at m/z 408, 410, and 412 (1:2:1), indicating the presence of two bromine atoms in the molecule, and the molecular formula was established as C₁₂H₁₉ON₅Br₂ by HRESIMS [m/z 408.0045 (M + H)⁺, Δ +1.0 mmu]. The UV absorption [λ_{max} 273 (ϵ 4500) nm]



Figure 1. Selected 2D NMR correlations for nagelamide O (1).



Figure 2. Selected NOESY correlations and relative stereochemistry of the tetracyclic core in nagelamide O (1).

was attributed to a substituted pyrrole chromophore, while IR absorptions indicated the occurrence of OH (3440 cm^{-1}) and amide carbonyl (1680 cm^{-1}) functionalities. ¹H and ¹³C NMR data (Table 3) of **3** suggested the presence of a dibromopyrrole carbonyl moiety (N-1–C-6) and a guanidino moiety (N-13, C-14, N-15, and N-16).⁹ Detailed analysis of the ¹H–¹H COSY spectrum disclosed the connectivity from H-8 to NH-13 (Figure 5). HMBC correlations for NMe-17/C-6 and NMe-17/C-8 and the NOESY correlation for H-4/NMe-17 indicated that a 2,3-dibromopyrrole carbonyl moiety was connected to N-7 through an amide bond. Thus, the structure of mukanadin E was assigned as **3**, which is the 2-bromo and 7-methyl analogue of laughine.⁹

The molecular formula of mukanadin F (4) was established as $C_{11}H_{10}O_4N_4Br_2$ by HRESIMS $[m/z 442.8967 (M + Na)^+, \Delta + 0.1]$ mmu]. The chemical shifts of C-10 ($\delta_{\rm C}$ 111.6), C-11 ($\delta_{\rm C}$ 130.5), C-12 ($\delta_{\rm C}$ 164.6), and C-14 ($\delta_{\rm C}$ 154.7) suggested the presence of a hydantoin ring.10 The 1H-1H COSY spectrum of 4 showed the connectivity from NH-7 to H-10, indicating that the hydroxyl group was attached to C-9 ($\delta_{\rm H}$ 4.43 and $\delta_{\rm C}$ 65.7). The connectivity of a dibromopyrrole ring to N-7 was elucidated from a NOESY correlation for H-4/NH-7, while HMBC cross-peaks of H-9/C-11, H-10/C-12, NH-13/C-11, NH-15/C-14, and NH-15/C-11 indicated that C-10 was connected to the hydantoin ring at C-11. The geometry of the double bond at C-10 was assigned as Z from the NOESY correlation for OH-9/NH-15. To analyze the absolute stereochemistry at C-9, (S)- and (R)-MTPA esters of mukanadin F (4) were prepared. $\Delta\delta$ values for (S)- and (R)-MTPA esters of mukanadin F (4) suggested the absolute configuration at C-9 to be R^{11} Thus, the structure of mukanadin F was elucidated as 4, which is the 9-hydroxy form of mukanadin D.10

Nagelamide O (1) is a rare bromopyrrole alkaloid with a perhydrocyclopenta-imidazo-azolo-imidazole carbon skeleton, while mukanadin E (3) is the first bromopyrrole alkaloid possessing an *N*-methyl amide bond. Nagelamide O (1) showed weak antibacterial activity against *Bacillus subtilis*, *Micrococcus luteus*, and *Staphylococcus aureus* (MIC, 33.3 μ g/mL each), while mukanadin F (4) exhibited weak antifungal activity against *Aspergillus niger* (MIC, 16.7 μ g/mL).¹²

Experimental Section

General Experimental Procedures. Optical rotations were recorded on a JASCO P-1030 polarimeter. UV spectra were recorded on a Shimadzu UV-1600PC spectrophotometer. CD spectra were recorded on a JASCO J-720 polarimeter. IR spectra were recorded on a JASCO FT/IR-230 spectrophotometer. ¹H and ¹³C NMR spectra were recorded on a Bruker AMX-600 and a JEOL ECA-500 spectrometer. The 2.49 and 39.5 ppm resonances of residual DMSO-*d*₆ were used as internal references for ¹H and ¹³C NMR spectra, respectively. ESIMS were obtained on a JEOL JMS-700TZ spectrometer.



Figure 3. CD (A) and UV (B) spectra, and rotation model for the C-9-C-9' bond of nagelamide O (1).

position	$\delta_{ m H}$	mult. J (Hz)	$\delta_{ m C}$	position	$\delta_{ m H}$	mult. J (Hz)	$\delta_{ m C}$
1	12.68	brs		1'	11.80	brs	
2			104.6	2'	6.98	brs	121.4
3			98.0	3'			95.0
4	6.95	brs	112.8	4'	6.86	brs	111.7
5			128.1	5'			126.6
6			159.5	6'			158.8
7	8.42	brt, 5.9		7'	8.39	brt, 5.9	
8	3.87	brt, 5.3	40.6	8'	3.95	m	40.5
9	6.02	dt, 16, 5.3	139.1	9'	6.02	dt, 16, 5.3	136.6
10	6.05	brd,	126.7	10'	6.41	brd, 16	115.7
11			69.9	11'			119.2
12				12'	12.56	brs	
13			147.7	13'			147.7
NH ₂ -13	7.62	brs, 2H		NH ₂ -13'	7.62	brs, 2H	
14	12.12	brs		14'			
15			183.4	15′			121.3

Table 2. ¹H and ¹³C NMR Data of Nagelamide P (2) in DMSO- d_6

Sponge Description. The yellowish-brown sponge *Agelas* sp. (SS-983, family Agelasidae) collected off Seragaki, Okinawa, and the redbrown sponge *Agelas* sp. (SS-1056) collected off Gesashi, Okinawa, and the sponge *Agelas* sp. (SS-1077) collected off Unten-Port, Okinawa, were kept frozen until used. The voucher specimens (SS-983, SS-1056, and SS-1077) were deposited at the Graduate School of Pharmaceutical Sciences, Hokkaido University.

Extraction and Isolation. The sponge SS-983 (0.2 kg, wet wt) was extracted with MeOH, the extract was partitioned between EtOAc and H₂O, and subsequently the aqueous layer was extracted with *n*-BuOH. *n*-BuOH-soluble materials were subjected to LH-20 column chromatography to give an alkaloid-containing fraction. This fraction was separated by a C₁₈ column chromatography (MeOH/H₂O/CF₃COOH, 30:70:0.1 \rightarrow 100:0:0) and then C₁₈ HPLC (YMC-Pack ODS-AM, YMC Co., Ltd., 10 × 250 mm; eluent CH₃CN/H₂O/CF₃COOH, 37.5:62.5:



Table 3. ¹H and ¹³C NMR Data of Mukanadins E (3) and F (4) in DMSO- d_6

	3		4		
position	$\delta_{ m H}$	$\delta_{ m C}$	$\delta_{ m H}$	$\delta_{ m C}$	
1	12.27 brs		12.67 brs		
2		103.57		104.56	
3		97.39		97.86	
4	6.60 brs	113.71	6.93 brs	112.92	
5		126.92		128.00	
6		160.05		159.05	
7			8.06 t, 5.4		
8	3.44 t	48.05	3.29 m	44.43	
9	1.58 m	27.76	4.43 m	65.67	
OH-9			5.34 m		
10	1.30 m	22.93	5.45 d, 8.4	111.59	
11	1.52 m	26.29		130.45	
12	3.12	40.43		164.64	
13	7.02		10.02 brs		
14		156.82		154.68	
15			11.02 brs		
16					
Me-17	3.08 s	35.09			

0.1; flow rate 2 mL/min; UV detection at 254 nm) to yield nagelamides O (1, 2.7 mg) and P (2, 1.0 mg). The sponge *Agelas* sp. (SS-1056, 1.35 kg, wet wt) was extracted with MeOH, and the extract was partitioned between EtOAc and H₂O. EtOAc-soluble materials were

Figure 4. Selected 2D NMR correlations for nagelamide P (2).



Figure 5. Selected 2D NMR correlations for mukanadin E (3).



Figure 6. Selected 2D NMR correlations for mukanadin F (4).

subjected to a silica gel column chromatography (CHCl₃/*n*-BuOH/ AcOH/H₂O, 1.5:6:1:1) to give an alkaloid-containing fraction. This fraction was separated by a C₁₈ column chromatography (MeOH/H₂O/ CF₃COOH, 80:20:0.1) and then C₁₈ HPLC (YMC-Pack ODS-AM, YMC Co., Ltd., 10 × 250 mm; eluent CH₃CN/H₂O/CF₃COOH, 25: 75:0.1 \rightarrow 35:65:0.1; flow rate 2 mL/min; UV detection at 255 nm) to yield mukanadin E (**3**, 4.3 mg). Mukanadin F (**4**, 6.6 mg) was obtained from the sponge *Agelas* sp. (SS-1077, 0.70 kg, wet wt) by the same procdure as for mukanadin E (**3**).

Nagelamide O (1): colorless, amorphous solid; $[α]_D^{22}$ +1.6 (*c* 0.75, MeOH); UV (MeOH) λ_{max} 275 (*ε* 9400) nm; IR (KBr) ν_{max} 3450, 2920, 2850, 1680, 1210 cm⁻¹; CD (MeOH) λ_{max} 281 (Δ*ε* +5.2) and 257 (Δ*ε* -3.4) nm; ¹H and ¹³C NMR data, see Table 1; ESIMS (positive) *m/z* 765:767:769:771:773 [(M + H)⁺, 3:10:12:6:1]; HRESIMS *m/z* 764.9315 [(M + H)⁺, Δ +1.6 mmu], calcd for C₂₂H₂₅⁷⁹Br₃³⁵ClN₁₀O₄, 764.9299.

Nagelamide P (2): colorless, amorphous solid; $[α]_D^{22} ~ ~0$ (*c* 0.5, MeOH); UV (MeOH) $λ_{max}$ 276 (ϵ 21 100) nm; IR (KBr) $ν_{max}$ 3350, 2920, 2850, 1680, 1210 cm⁻¹; ¹H and ¹³C NMR data, see Table 2; ESIMS *m*/*z* 711:713:715:717 [(M + H)⁺, 1:3:3:1]; HRESIMS *m*/*z* 710.9465 [(M + H)⁺, Δ +3.8 mmu], calcd for C₂₂H₂₁⁷⁹Br₃N₁₀O₃, 710.9427.

Mukanadin E (3): colorless, amorphous solid; UV (MeOH) λ_{max} 273 (ϵ 4500) nm; IR (KBr) ν_{max} 3440, 1680 cm⁻¹; ¹H and ¹³C NMR, see Table 3; ESIMS *m*/*z* 408, 410, and 412 [(M + H)⁺, 1:2:1]; HRESIMS *m*/*z* 408.0045 [(M + H)⁺, Δ +1.0 mmu], calcd for C₁₂H₂₀ON₅⁷⁹Br₂, 408.0035.

Mukanadin F (4): colorless, amorphous solid; $[α]_D^{20} - 3.9$ (*c* 0.3, MeOH); UV (MeOH) λ_{max} 279 (ϵ 16 000), 203 (32 000) nm; IR (KBr) ν_{max} 3210, 1685 cm⁻¹; ¹H and ¹³C NMR, see Table 3; ESIMS *m/z* 443, 445, and 447 [(M + Na)⁺, 1:2:1]; HRESIMS *m/z* 442.8967 [(M + Na)⁺, Δ +0.1 mmu], calcd for C₁₁H₁₀O₄N₄⁷⁹Br₂ Na, 442.8966.

Preparation of (*R***)- and (***S***)-MTPA Esters of Mukanadin F (4).** To a solution of **4** (0.4 mg) in pyridine (100 μ L) were added *N*,*N*-demethylaminopyridine (100 μ g), triethylamine (6 μ L), and (*S*)-MTPA Cl (3 μ L), and stirring was continued at room temperature for 24 h. After removal of solvent, resulting residue was partitioned between EtOAc (200 μ L × 3) and H₂O (100 μ L × 3). The organic layer was concentrated *in vacuo* to afford the (*R*)-MTPA ester of **4**. The (*S*)-MTPA ester of **4** was prepared according to the same procedure as described above.

Acknowledgment. We thank Mr. Z. Nagahama, Mr. S. Furugen, and Mr. K. Uehara for their help with collection of the sponge, and Ms. S. Oka, Center for Instrumental Analysis, Hokkaido University, for the ESIMS measurements. This work was partly supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology of Japan.

References and Notes

- Blunt, J. W.; Copp, B. R.; Hu, W.-P.; Munro, M. H. G.; Northcote, P. T.; Princep, M. R. Nat. Prod. Rep. 2008, 25, 35–94.
- (2) Takahashi, Y.; Kubota, T.; Ito, J.; Mikami, Y.; Fromont, J.; Kobayashi, J. Bioorg. Med. Chem. 2008, 16, 7561–7564.
- (3) Araki, A.; Kubota, T.; Tsuda, M.; Mikami, Y.; Fromont, J.; Kobayashi, J. Org. Lett. 2008, 10, 2099–2102.
- (4) Kubota, T.; Araki, A.; Ito, J.; Mikami, Y.; Fromont, J.; Kobayashi, J. *Tetrahedron* **2008**, *64*, 10810–10813.
- (5) Urban, S.; Leon, P.de A.; Carroll, A. R.; Fechner, G. A.; Smith, J.; Hooper, J. N. A.; Quinn, R. J. J. Org. Chem. 1999, 64, 731–735.
- (6) Nishimura, S.; Matsunaga, S.; Shibazaki, M.; Suzuki, K.; Furihata, K.; van Soest, R. W. M.; Fusetani, N. Org. Lett. 2003, 5, 2255–2257.
- (7) Grube, A.; Immel, S.; Baran, P. S.; Köck, M. Angew. Chem., Int. Ed. 2007, 46, 6721–6724.
- (8) Tsukamoto, S.; Kato, H.; Hirota, H.; Fusetani, N. J. Nat. Prod. 1996, 59, 501–503.
- (9) Williams, D. E.; Patrick, B. O.; Behrisch, H. W.; van Soest, R. W. M.; Roberge, M.; Andersen, R. J. J. Nat. Prod. 2005, 68, 327–330.
- (10) Hu, J. F.; Peng, J. N.; Kazi, A. B.; Kelly, M.; Hamann, M. T. J. Chem. Res. 2005, 7, 427–428.
- (11) Ohtani, I.; Kusumi, T.; Kashman, Y.; Kakisawa, H. J. Am. Chem. Soc. **1991**, 113, 4092–4095.
- (12) Nagai, H.; Mikami, M.; Yazawa, K.; Gonoi, T.; Yasumoto, T. J. Antibiot. 1993, 46, 520–522.

NP800645Q